## The Total Synthesis of Antrimycin D<sub>v</sub>

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Antrimycin  $D_{v}$ , a representative of the antrimycin and cirratiomycin classes of tuberculostatic heptapeptide antibiotics containing a tetrahydropyridazinecarboxylic acid unit, has been synthesized.

The closely related (and, in parts, identical) cirratiomycins<sup>1</sup> **1** and antrimycins<sup>2</sup> **2** have been isolated from *Streptomyces xanthocidicus* and *Streptomyces cirratus*. These tuberculostatic peptides contain the non-proteinogenic amino acids  $\beta$ , $\beta$ -dihydroxy- $\alpha$ -aminoisobutyric acid, didehydrovaline or didehydroisoleucine, (*S*,*S'*)- $\alpha$ , $\beta$ -diaminobutyric acid and (*S*)tetrahydropyridazinecarboxylic acid. The latter has now been detected for the first time whereas its 4-hydroxy derivative had been found as a residue of luzopeptin,<sup>3</sup> a DNA intercalating, cancerostatic cyclopeptide.

The construction of the completely protected  $\beta$ , $\beta$ -dihydroxy- $\alpha$ -aminoisobutyric acid is shown in Scheme 1. The nitro alcohol **3** is readily accessible from tris(hydroxymethyl)nitromethane and was converted to the Boc-protected amino acid **4a** by a two-step oxidation; **4a** can be activated as its pentafluorophenyl ester **4b**.

The masked didehydrotripeptide Boc-Leu-D,D-Val-Ser-OAll 7 was prepared as shown in Scheme 2. The didehydrodipeptide 6 was obtained from the dipeptide 5 by a Horner condensation. Analogous reactions have been reported frequently.<sup>4</sup> When DBU is employed as the base, ketones can also take part in the phosphonic ester condensation.<sup>5</sup>

We have developed two routes for the preparation of the 2,3,4,5-tetrahydropyridazinecarboxylic acid in which the ring is built up through intramolecular hydrazone formation from a 2-hydrazino-4-formylbutyric acid (Scheme 3). The protected 4-formylbutyric acid derivative 9 is metallated and converted to the hydrazine 10 by reaction with an azodicarboxylate according to Evans's method. The latter is converted to the cyclic hydrazone 12 by opening of the cyclic ketal and cleavage of the Boc group. Unfortunately, the free ester is unstable and cannot be acylated.

Model experiments, however, have shown that N<sup> $\alpha$ </sup>-acylated hydrazines (**11**, N<sup> $\alpha$ </sup>-Ac in place of N<sup> $\alpha$ </sup>-Boc) form stable *N*-acyl derivatives of tetrahydropyridazinecarboxylic acid in trifluoroacetic acid. This sequence was used to construct the protected diaminobutyryltetrahydropyridazinecarboxylate **15**. A suitably protected diaminobutyric acid derivative was available from a previous synthesis of lavendomycin.<sup>6</sup> The protected 4-formyl-2-oxobutyrate **13** was converted to the Boc-hydrazone which, in turn, was reduced to the hydrazine derivative **14**. Coupling with the  $\alpha$ , $\beta$ -diaminobutyric acid



derivative and acid treatment gave rise to the dipeptide 15. Subsequent cleavage of the Fmoc group, coupling with Boc-alanine to give 16, and reduction of the azide group furnished the tripeptide 17 as a mixture of diastereoisomers (1:1). The diastereoisomers can be separated easily by MPLC at the stages of 16 and 17a. The assignment was deduced later from a comparison of the NMR spectra of the diastereoisomeric hexapeptides with that of the naturally occurring product. Only in the case of one of the diastereoisomers 18 and *epi*-18 were the chemical shifts of the signals of the two methyl groups of D,D-valine in agreement with those of the natural product.

The 'correct' diastereoisomer of **16b** was converted to the hexapeptide **18** by coupling with the dehydrotripeptide **7b**. Cleavage of the Boc group and acylation with the hydroxymethylserine derivative **4b** gave rise to the completely



Boc = *tert*-butoxycarbonyl

Scheme 1 Reagents and conditions: i, H<sub>2</sub>, Pd/C, EtOH, room temp., 3 days, 98%; ii, Boc<sub>2</sub>O, CH<sub>2</sub>Cl<sub>2</sub>, room temp., 14 h, 99%; iii, (COCl)<sub>2</sub>, dimethyl sulfoxide (DMSO), CH<sub>2</sub>Cl<sub>2</sub>, -60 °C, 10 min, triethylamine (TEA), -60 °C, 15 min, -60 °C to room temp. 1 h, 95%; iv, NH<sub>2</sub>SO<sub>3</sub>H, NaHPO<sub>4</sub>, NaClO<sub>2</sub>, H<sub>2</sub>O-dioxane, 5 °C, 30 min, room temp., 30 min, 72%; v, C<sub>6</sub>F<sub>5</sub>OH, dicyclohexylcarbodiimide (DCC), CH<sub>2</sub>Cl<sub>2</sub>, -20 to 20 °C, 16 h, 100%



Scheme 2 Reagents and conditions: i, 1,8-diazabicyclo[5.4.0]undec-7ene (DBU), acetone, room temp., 48 h, 75%; ii, NaOH, H<sub>2</sub>Odioxane, room temp., 16 h, 83%; iii, N-(3-dimethylaminopropyl)-N'ethylcarbodiimide hydrochloride (EDC), Et<sub>3</sub>N, hydroxybenzotriazole, dimethylformamide (DMF), H-Ser-OAll·xHCl, -20 °C to room temp., 16 h, 52%; iv, 6 mol dm<sup>-3</sup> HCl-dioxane, 5 to 20 °C, 2 h, 100%



*Abbreviations*: Fmoc = fluoren-9-ylmethoxycarbonyl; Alloc = allyloxy-carbonyl; All = allyl; Succ = succinimido

Scheme 3 Reagents and conditions: i, NaOH, H<sub>2</sub>O-dioxane, room temp., 14 h, 93%; ii, 1-chloro-N,N,2-trimethylprop-1-en-1-amine, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, 10 min, lithium salt of (4*S*)-4-benzyl-2-oxazolidinone, tetrahydrofuran (THF), -80 °C, 15 min, -80 to 0 °C, 2 h, 76%; iii, NaN(SiMe<sub>3</sub>)<sub>2</sub>, THF, -80 °C, 30 min, di-*tert*-butoxycarbonyl azocarboxylate, CH<sub>2</sub>Cl<sub>2</sub>-THF, 3 min, 67%; iv, LiOH, H<sub>2</sub>O-THF, 0 °C, 2 h, v, CH<sub>2</sub>N<sub>2</sub>, Ch<sub>2</sub>Cl<sub>2</sub>, room temp., 2 min, iv + v: 99%; vi, CF<sub>3</sub>CO<sub>2</sub>H, room temp., 15 min, 90%; vii, Boc-NHNH<sub>2</sub>, hexane, reflux, 3 h, 64%, viii, NaOH, H<sub>2</sub>O-dioxane, room temp., 4 h, 86%; ix, CH<sub>2</sub>N<sub>2</sub>,

protected antrimycin  $D_v$  **19**.<sup>†</sup> The two allyl protecting groups were cleaved by<sup>7</sup> Pd<sup>0</sup> while the protecting groups on the hydroxymethylserine unit were removed by acid treatment. The thus synthesized compound was identical with the natural product.

Support of this research work by BASF AG, the Fonds der Chemischen Industrie, the Deutsche Forschungsgemeinschaft and the Land Baden-Württemberg is gratefully acknowledged. We thank Prof. G. Jung and Dr. J. Metzger (Universität Tübingen) for mass spectra and cand. chem. Jürgen Heim and cand. chem. Heinz Greisiger for their help and encouragement.

Received, 13th April 1992; Com. 2/01926D

## References

- Isolation, structure and biological activities: T. Shiroza, N. Ebisawa, A. Kojima, K. Furihata, A. Shimazu, T. Endo, H. Seto and N. Otake, Agric. Biol. Chem., 1982, 46, 1885; T. Shiroza, N. Ebisawa, K. Furihata, T. Endo, H. Seto and N. Otake, Agric. Biol. Chem., 1982, 46, 1891; T. Shiroza, N. Ebisawa, K. Furihata, T. Endo, H. Seto and N. Otake, Agric. Biol. Chem., 1982, 46, 865.
- Isolation and structure: N. Shimada, K. Morimoto, H. Naganawa, T. Takita, M. Hamada, K. Maeda, T. Takeuchi and H. Umezawa, J. Antibiot., 1981, 34, 1613; H. Morimoto, N. Shimada, H. Naganawa, T. Takita and H. Umezawa, J. Antibiot., 1981, 34, 1615; H. Morimoto, N. Shimada, H. Naganawa, T. Takita and H. Umezawa, J. Antibiot., 1982, 35, 378.
- 3 Isolation, structure and biological activities: M. Konishi, H. Ohkuma, F. Sakai, T. Tsuno, H. Koshiyama, T. Naito and H. Kawaguchi, J. Am. Chem. Soc., 1981, 103, 1241; E. Arnold and J. Clardy, J. Am. Chem. Soc., 1981, 103, 1243.
- 4 U. Schmidt, A. Lieberknecht and J. Wild, Synthesis, 1988, 159; U. Schmidt and J. Wild, Liebigs Ann. Chem., 1985, 1882.
- Schmidt and J. Wild, *Liebigs Ann. Chem.*, 1985, 1882.
  U. Schmidt, H. Griesser, V. Leitenberger, A. Lieberknecht, R. Mangold, R. Meyer and B. Riedl, *Synthesis*, 1992, 487.
- 6 U. Schmidt, K. Mundinger, R. Mangold and A. Lieberknecht, J. Chem. Soc., Chem. Commun., 1990, 1216.
- 7 P. P. Jeffrey and S. W. McCombie, J. Org. Chem., 1982, 47, 587;
   H. Kunz and H. Waldmann, Angew. Chem., 1984, 96, 49.

† <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta$  8.22 (d, *J* 6.7 Hz, 1H), 8.17 (s, 1H), 7.35–7.51 (m, 7H), 7.23 (d, *J* 6.7 Hz, 1H), 7.09 (s, 1H), 5.82–5.98 (m, 2H), 5.60–5.63 (m, 1H), 5.57 (s, 1H), 5.16–5.38 (m, 4H), 5.0 (s, 1H), 4.59–4.68 (m, 4H), 4.42–4.52 (m, 2H), 4.22–4.39 (m, 5H), 3.84–4.02 (m, 2H), 3.37–3.72 (m, 1H), 2.17–2.23 (m, 2H), 2.11 (s, 3H), 1.80–1.85 (m, 1H), 1.76 (s, 3H), 1.55–1.71 (m, 4H), 1.50 (s, 9H), 1.35 (d, *J* 7.0 Hz, 3H), 1.09 (d, *J* 6.8 Hz, 3H), 0.94 (d, *J* 6.7 Hz, 3H), 0.90 (d, *J* 5.5 Hz, 3H).

CH<sub>2</sub>Cl<sub>2</sub>, room temp., 5 min, 98%; x, NaCNBH<sub>3</sub>, HOAc, MeCN, room temp., 24 h, 98%; xi, Fmoc-NH-CH(CHMeN<sub>3</sub>)COCl, collidine, THF, 0 °C, 2 h, room temp., 16 h; xii, CF<sub>3</sub>CO<sub>2</sub>H, room temp., 2 h, xi + xii: 52%; xiii, NaOH, H<sub>2</sub>O-dioxane, room temp., 14 h, xiv, H<sub>2</sub>SO<sub>4</sub>, dioxane–H<sub>2</sub>O, room temp., 15 min, xv, NaHCO<sub>3</sub>, Boc-Ala-OSuc/, H<sub>2</sub>O-dioxane, room temp., 16 h, xii + xiv + xv: 74%; xvi, H<sub>2</sub>, Pd/C, dioxane, room temp., 3 h, xvii, Alloc-Cl, pyridine, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, 1 h, room temp., 14 h, xvi + xvii: 84%; xviii, NaOH, H<sub>2</sub>O-dioxane, room temp., 14 h, 100%; xix, 7b, EtNPri<sub>2</sub>, diphenyl phosphorylazide, CH<sub>2</sub>Cl<sub>2</sub>, -20°C, 1 h, 4°C, 3 days, 68%; xx, 6 mol dm<sup>-3</sup> HCl-dioxane, 5 to 20 °C, 2 h, 100%; xxii, 4b, dimethylaminopyridine (DMAP), THF, room temp., 16 h, 47%; xxii, Pd(PPh<sub>3</sub>)<sub>4</sub>, morpholine, THF, room temp., 2 h, xxiii, 6 mol dm<sup>-3</sup> HCl-H<sub>2</sub>O, room temp., 2 h, xxiii + xxiii: 56%